# 43. Synthesis of a Phosphonic Acid Analogue of N-Acetyl-2,3-didehydro-2deoxyneuraminic Acid, an Inhibitor of Vibrio cholerae Sialidase

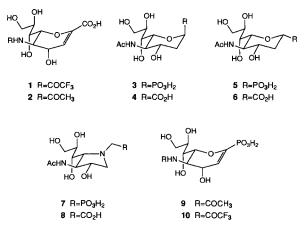
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## (25.I.91)

The synthesis of the phospha analogue 10 of DANA (2) is described. Bromo-hydroxylation of the known 11  $(\rightarrow 12 \text{ and } 13)$  followed by treatment of the major bromohydrin 13 with 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) gave the oxirane 14 (*Scheme 1*). Depending on the solvent, TiBr<sub>4</sub> transformed 14 into 16 or into a 15/16 mixture. Reductive debromination of 16  $(\rightarrow 17)$ , followed by benzylation provided 18. Oxidative decarboxylation (Pb(OAc)<sub>4</sub>) of the acid, obtained by saponification of 18, yielded the anomeric acetates 19 and 20. While 19 was inert under the conditions of phosphonoylation, the more reactive imidate 22, obtained together with 23 from 19/20 via 21 (*Scheme 2*), gave a mixture of the phosphonates 27/28 since  $\beta$ -elimination of AcOH from 27/28 proved difficult, the bromide 34 was prepared from 27/28 by photobromination and subjected to reductive elimination with Zn/Cu ( $\rightarrow$ 35; *Scheme 3*). This two-step sequence was first investigated using the model compounds 30 and 31. Transesterification of 35, followed by deacetylation gave 10, which is a strong inhibitor of the *Vibrio Cholerae* sialidase.

**Introduction.** – FANA (2,6-anhydro-3,5-dideoxy-5-(trifluoroacetamido)-D-glycero-Dgalacto-non-2-enonic acid; 1) and DANA [1] (5-acetamido-2,6-anhydro-3,5-dideoxy-Dglycero-D-galacto-non-2-enonic acid; 2) are among the strongest inhibitors of *N*acetylneuraminidases of different origin. We have described the three phosphonates 3, 5, and 7 which are all stronger inhibitors of *Vibrio cholerae* neuraminidase than the corresponding carboxylates 4, 6, and 8 [2][3]. The  $K_i$  values of 3 and 5 are  $2.3 \cdot 10^{-4}$  M and  $7.5 \cdot 10^{-5}$  M, respectively, as compared to  $2.6 \cdot 10^{-3}$  M for 6, while 4 is inactive. We were, therefore, interested in the inhibitory effect of the phospha analogues 9 and 10 of FANA (1) and DANA (2).



As we had prepared 2 by  $\beta$ -elimination of a suitably protected  $\beta$ -acetoxy ester [4][5], we investigated an analogous route to 10 which should also be feasable for the preparation of 9. The preparation of 10 then involves introduction of a leaving group at C(3), decarboxylation, and phosphonoylation. As phosphonoylation [6] is expected to proceed better in the presence of *O*-alkyl instead of *O*-acyl protective groups [7], we planned to first prepare the benzylated acetates 19/20 (*Scheme 1*). For this, we thought to use the previously developed halogenohydroxylation [4][8] of Neu2en5Ac derivatives such as 11 to refunctionalize C(3) and a decarboxylation by Pb(OAc)<sub>4</sub> [2]. Syntheses of 1-alkenyl phosphonates by  $\beta$ -elimination are known [9].

**Results and Discussion.** – Bromohydroxylation of 11<sup>1</sup>) with *N*-bromosuccinimide (NBS) and H<sub>2</sub>O in MeCN at 70° [10] gave the bromohydrins 12 and 13 (98%; 12/13 1:2.8, *Scheme 1*)<sup>2</sup>). Treatment of 13 with 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) [8] gave the epoxide 14 (85%). Br–C(3) is axial in 13 (J(3,4) = 3.5 Hz) and equatorial in 12 (J(3,4) = 9.7 Hz). The long-range W-coupling  ${}^{4}J(3,OH-C(2)) = 1.2$  Hz observed in the <sup>1</sup>H-NMR spectrum of 12 also proves the axial orientation of OH–C(2), since the requirements for such a coupling are not fulfilled by an equatorial OH–C(2) [11]. The <sup>1</sup>H-NMR spectrum of 14 shows a *s* for H–C(3) at 3.64 ppm (for similar examples, see [8]), indicating a dihedral angle between H–C(3) and H–C(4) of *ca.* 90° and thus a  ${}^{5}H_{4}$  or a  ${}^{6}S$  conformation of the pyranose ring.

The reaction of the epoxide 14 with  $TiBr_4$  gave a mixture of the desired bromohydrin 16 and of the bicyclic acetal 15 (95%; 16/15 1.8:1), when the reaction was carried out in 1,2-dichloroethane at  $-78^{\circ}$ , but exclusively the desired 16 in THF at  $-40^{\circ}$  (96%). The 2,7-anhydro derivative 15 must be formed by interception of the (incipient) intermediate carboxonium ion by the BnO–C(7) group<sup>3</sup>). The suppression of this side reaction in THF may be due to an intermolecular solvation of the carboxonium ion by THF competing successfully with BnO–C(7). Reductive debromination of 16 with tributyltin hydride (Bu<sub>3</sub>SnH) gave the alcohol 17 (91%).

The IR spectrum of **15** shows an OH band at 3460 cm<sup>-1</sup>, and the <sup>13</sup>C- and <sup>1</sup>H-NMR spectra show the presence of only 3 BnO groups. A downfield shift of 1–2 ppm for the NH signal (as compared to **12–14** and **16–18**) evidences the presence of an H-bond NH···O–C(3). The small vicinal coupling constants for the pyranose-ring protons (< 2 Hz) and the long-range couplings J(3,5) and J(4,6) (1.6 and 1.4 Hz, resp.) indicate a  ${}^{5}C_{2}$  conformation, compatible with a 6,8-dioxa[3.2.1]bicyclooctane structure. In the <sup>1</sup>H-NMR spectra of **17**, H–C(2) resonates at 3.63 ppm, with J(2,3) = 9.6 Hz proving its axial orientation. The exclusive axial attack of Bu<sub>3</sub>SnH is in agreement with our previous results [13][4].

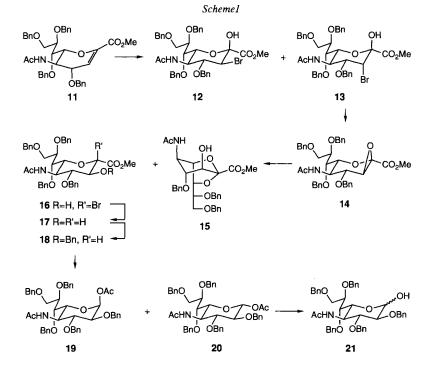
Benzylation of 17 (PhCH<sub>2</sub>Br, NaH, DMF) yielded the penta-O-benzyl derivative 18 (63%) together with unreacted 17 (26%). Hydrolysis of 18 gave the corresponding acid which was oxidatively decarboxylated with Pb(OAc)<sub>4</sub> [14] in toluene and in the presence of pyridine<sup>4</sup>) to a mixture of the anomeric acetates 19 and 20 (*Scheme 1*; 74%; 19/20

<sup>&</sup>lt;sup>1</sup>) Obtained in five steps from *N*-acetylneuraminic acid (Neu5Ac) according to known procedures [3].

<sup>&</sup>lt;sup>2</sup>) Ito and Ogawa indicate a ratio 12/13 of 1:4.1 [10].

<sup>&</sup>lt;sup>3</sup>) Such a neighbouring group participation [12] and particularly the formation of 6,8-dioxa[3.2.1]bicyclooctane structures similar to **15** have been described in some detail; see [5] and refs. cit. therein.

<sup>&</sup>lt;sup>4</sup>) Almost no reaction occurred in the absence of pyridine.



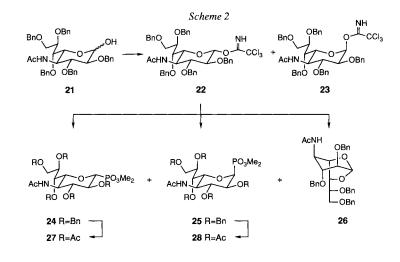
 $Bn = PhCH_2$ 

3.3:1). The anomeric configuration of **19** and **20** is deduced from the chemical shifts of H–C(1) and  ${}^{3}J(1,2)$  (**19**: 6.33 ppm, J = 3.5 Hz; **20**: 5.57 ppm, J = 8.2 Hz; *Table 1*). The specific rotations of **19** and **20** confirm this assignment (**19**:  $[\alpha]_{D} = -27.4$ , and **20**:  $[\alpha]_{D} = -3.4$ ). The acetate **19** proved inert to trimethyl phosphite (P(OMe)<sub>3</sub>) in the presence of trimethylsilyl triflate (Me<sub>3</sub>SiOTf) [2][15].

	H <sub>a</sub> a)	J(a,b)	[ <i>M</i> ] <sup>25</sup> <sub>D</sub>	Anomeric configuration		
17	3.63	9.6	-65.0			
18	3.80	<sup>b</sup> )	-49.5			
19	6.33	3.5	-212.1	<b>β</b> -D		
20	5.57	8.2	-26.3	α-D		
22	5.78	8.3	-47.3	α-D		
23	6.51	3.2	-191.0	<b>β</b> -D		

Table 1. Selected Data of the Carboxylates 17 and 18, the Acetates 19 and 20, and the Imidates 22 and 23

Trichloroacetimidates are more reactive starting materials for the preparation of glycosylphosphonates than the corresponding acetates [7]. We, therefore, deacetylated the mixture 19/20 (NaOMe/MeOH) to obtain 21 (100%) as a 1:1.9 mixture of the  $\alpha$ - and the  $\beta$ -D-anomers<sup>5</sup>). Reaction of **21** with trichloracetonitrile and K<sub>2</sub>CO<sub>3</sub> in MeCN [17] gave the stable imidates 22 and 23 (70%; 22/23 2.4:1; Scheme 2) and unreacted 21 (14%). The IR spectra of 22 and 23 show the amide N-H bands at 3430 and the typical imide N-H bands at 3340 (22) and 3350  $\text{cm}^{-1}(23)$ . Similarly as for 19 and 20, the anomeric configuration of 22 and 23 was deduced from a comparison of the H–C(1) resonances (22: 5.78 ppm, J = 8.3Hz; and 23: 6.51 ppm, J = 3.2 Hz; Table 1), and from their specific rotations (22:  $[\alpha]_{\rm D} =$ -5.4; 23:  $[\alpha]_{\rm D} = -21.8$ ). Treating the imidate 22<sup>6</sup>) with P(OMe)<sub>3</sub> and Me<sub>3</sub>SiOTf [15] under rigorously anhydrous conditions gave a mixture of the anomeric phosphonates 24 and 25 (44%; 24/25 ca. 1:1) and of the bicyclic derivative 26 (41%), formed in a similar way as 15. Debenzylation of the mixture 24/25 followed by acetylation gave the pentaacetates 27 and 28 (97%). The equatorial 1,2-trans-phosphonate 24 was obtained in a surprisingly high proportion, considering both the results of Meuwly and Vasella [15] who found that mild reaction conditions lead mainly to the 1,2-cis-phosphonate and those of Vaghefi et al. [16] and of Briner and Vasella [6][7] who found that harsher conditions lead mostly to axial phosphonates. The results of Meuwly and Vasella [15] have been rationalized on the basis of a stabilizing coordination between the P-centre in the phosphonium-salt intermediate and a cis-oriented neighbouring BnO group [15], and this rationalization can also be applied to the present case. Formation of a six-membered ring by coordination between the P-centre and BnO-C(6) in the phosphonium-salt intermediate may indeed compete with the formation of a four-membered ring by coordination with BnO-C(2), and this may lead to a mixture 24/25.

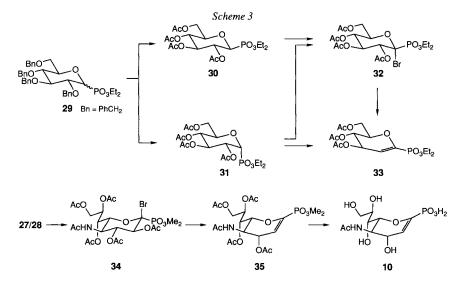


#### Bn = PhCH<sub>2</sub>

<sup>&</sup>lt;sup>5</sup>) The ratio of the anomers was determined by <sup>1</sup>H-NMR spectroscopy.

<sup>&</sup>lt;sup>6</sup>) Under similar conditions, the imidate 23 did not react.

Preliminary experiments showed that  $\beta$ -elimination from 27/28 was not straightforward. We, therefore, examined the  $\beta$ -elimination from the model compound 31, which was obtained in excellent yields by debenzylation and acetylation of 29 [15], together with a small amount of 30 (*Scheme 3*). As several conditions led to the elimination of AcOH, but not to completion of the reaction, we investigated an alternative reaction sequence. Photobromination according to a procedure by *Ferrier* and coworkers [18] (Br<sub>2</sub> or NBS in CCl<sub>4</sub>) gave exclusively the bromide 32 from either 30 or 31 (50–65%). Best results (76% of 33) for the reductive elimination from 32 were obtained with Zn/Cu in EtOH<sup>7</sup>) [19]. Similarly, photobromination of 27/28 yielded the bromide 34 (45%) together with unreacted 27/28 (15%). The acetoxy bromide 34 was converted into the vinylphosphonate 35 by treatment with activated Zn/Cu in EtOH (72%).



Similarly to **15**, the presence of an H-bond between N*H* and O–C(2) of **26** is deduced from the downfield shift of the N*H* signal (5.85 ppm as compared to 4.3–4.6 ppm for **19–24**). The presence of the dimethoxy-phosphono group in **24**, **25**, **27**, and in **28** is evidenced by the presence of 2 *d* for the MeO groups at 3.65–3.85 ppm (<sup>2</sup>*J*(C,P) = 10.6–10.9 Hz) in the <sup>1</sup>H-NMR spectra. The <sup>31</sup>P-NMR signals (see *Table 2*) show the presence of the phosphonate group for **24**, **25**, **27**, **28**, and **30** and **31**. Comparison of the <sup>31</sup>P-NMR shifts, the H–C(1) shifts, <sup>3</sup>*J*(H–C(2),P) and of <sup>1</sup>*J*(C(1),P) (*Table 2*) of **24/25**, **27/28**, and **30/31** allow the assignment of their anomeric configuration. As shown for the specific rotations of **27** ( $[\alpha]_D = -36.8$ ) and **28** ( $[\alpha]_D = +9.7$ ), *Hudson*'s rule [22] is followed.

On the basis of the low value of  ${}^{3}J(H-C(2),P)$  (6.2 and 6.5 Hz, *Table 2*), we assume that the newly introduced Br-atom in **32** and **34** is axial. The presence of a Br-substituent at C(1) is evidenced by two peaks of equal intensity for  $[M + 1]^+$  at m/z 549 and 547 for **32** and at m/z 664 and 662 for **34** and by the absence of the H–C(1) signal in the  ${}^{1}H$ -NMR spectrum. As photobromination proceeds by a radical mechanism, it is not surprising that only the axial bromide is found, regardless of the anomeric configuration of the starting material [18]. Photobromination occurred selectively at C(1), in keeping with the directing effect of the phosphonoyl group, whereas *Ferrier* and coworkers [18] observed mainly bromination at C(5) of  $\beta$ - and to a lesser extent also of  $\alpha$ -phexosides. The double bond of **33** and **35** was evidenced by the resonance of the H–C(2) signal at 5.80 and 5.79 ppm, respectively, and by the chemical shift of the C(1) (147.28 and 146.81 ppm) and C(2) signals (113.56 and 110.78 ppm).

<sup>7</sup>) Zn/Cu in AcOH/H<sub>2</sub>O gave 33 together with 30 and 31 [20][21].

	$\delta$ (H–C(1))	$2_{J(H-C(1),P)}$	$^{3}J(H-C(2),P)$	${}^{1}J(C(1),P)$	δ( <sup>31</sup> Ρ
24	< 3.90	b)	b)	с)	22.60
25	4.51	12.0	23.1	c)	23.99
27	3.75	10.4	9.2	173.2	18.60
28	4.74	10.2	30.8	152.4	21.56
30	3.83	10.2	10.3	172.7	16.18
31	4.66	11.4	29.6	154.0	19.74
32	-	_	6.2	195.6	8.06
33	_	_	10.5	225.2	6.70
34		_	6.5	195.5	9.46
35	-	-	10.6	228.5	9.25
10 <sup>d</sup> )	_	_	9.7	212.6	3.55

Table 2. Selected NMR Data of the Phosphonates 24, 25, 27, 28, 30-35, and 10a)

Table 3. <sup>1</sup>H,<sup>1</sup>H-Coupling Constants (Hz) of Neu5Ac, DANA (2), and 10<sup>a</sup>)

	J(3,4)	<i>J</i> (4,5)	J(5,6)	J(6,7)	J(7,8)	J(8,9)	J(8,9')	J(9,9')
Neu5Ac	11.8/5.0	10.4	10.7	1.2	9.4	2.8	6.4	-12.4
DANA (2)	2.5	8.9	10.9	1.2	9.3	2.7	6.0	-11.9
	J(2,3)	J(3,4)	J(4,5)	J(5,6)	J(6,7)	J(7,8)	J(7,8')	J(8,8')
10	2.2	8.9	10.7	_	9.8	2.6	6.5	-12.3

The phosphonate 35 was deprotected by transesterification with Me<sub>3</sub>SiBr [23], hydrolysis of the silyl ester, and deacetylation with NaOMe/MeOH to yield 54% of 10. Comparison of the coupling constants found in the <sup>1</sup>H-NMR spectra of 1, 2, and 10 indicates that these compounds possess identical conformations of the trihydroxypropyl side chain and of the dihydropyran ring (10 and 2; *Table 3*). The relatively low  $\delta$  (<sup>31</sup>P) values, and the relatively high  ${}^{1}J(C(1),P)$  values of the phosphonates 34 and 35 and of the phosphonic acid 10 (Table 2) are in agreement with earlier observations. Thus, alkyl, alkenyl, and alkynyl phosphonates on the one hand, and alkyl, 1-oxyalkyl, and (1-bromo-1-oxyalkyl) phosphonates on the other hand show decreasing values in this order for their <sup>31</sup>P-NMR shifts and increasing values for  ${}^{1}J(C(1),P)$  [24].

Inhibition of the Vibrio cholerae Sialidase by the Vinylphosphonic Acid 10. The phosphonic acid 10 was found to be a strong inhibitor of the Vibrio cholerae sialidase. The activity of the sialidase was reduced by 45, 62, and 80% at inhibitor concentrations of 0.1, 0.2, and 0.5 mm, respectively. The  $K_i$  value of 10 (7.2  $\cdot$  10<sup>-5</sup> m) was found to be slightly lower than for the anomeric phosphonates 3 and 5. The expected strong increase of the inhibitory power of the phosphonic acid 10 as compared to the corresponding carboxylic acid 2 was, however, not observed.

We thank the Swiss National Science Foundation and F. Hoffmann-La-Roche AG, Basle, for generous support.

### Experimental Part

General. See [25]. Methods for the sialidase experiments, see [5].

Methyl 5-Acetamido-4,7,8,9-tetra-O-benzyl-3-bromo-3,5-dideoxy- $\beta$ -D-erythro-L-gluco-2-nonulopyranosonate (12) and Methyl 5-Acetamido-4,7,8,9-tetra-O-benzyl-3-bromo-3,5-dideoxy- $\beta$ -D-erythro-L-manno-2-nonulopyranosonate (13). A soln. of 11 (11.5 g, 17.27 mmol) in MeCN (400 ml) and H<sub>2</sub>O (140 ml) was heated to 70°, and NBS (3.69 g, 20.76 mmol, 1.2 equiv.) was added. After stirring for 20 min., the soln. was cooled and evaporated. The residue was purified by chromatography (SiO<sub>2</sub>, AcOEt/hexane 1:4 to 2:1): 12 (3.373 g, 26%) and 13 (9.532 g, 72%).

Data of **12**. M.p. 153–154° (from Et<sub>2</sub>O/hexane) ([10]: 148–150°).  $R_f$  (AcOEt/hexane 1:1) 0.54.  $[\alpha]_D^{25} = -54.3$  (c = 1.11, CHCl<sub>3</sub>). IR (CHCl<sub>3</sub>): 3510m, 3440m, 3000m, 2960m, 2870m, 1750s, 1690s, 1510m, 1500m, 1455m, 1370m, 1280m, 1160s, 1120s, 1095s, 1050s, 1030s. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): 7.4–7.2 (m, 20 arom. H); 4.88 (d, J = 10.8, PhCH); 4.81 (d, J = 9.0, NH); 4.65 (d, J = 10.9, PhCH); 4.64 (d, J = 11.4, PhCH); 4.60–4.40 (m, 3 H, PhCH); 4.50 (m, H–C(6)); 4.50 (d, J = 10.9, PhCH); 4.64 (d, J = 11.3, PhCH); 4.50 (m, H–C(6)); 4.50 (d, J = 10.9, PhCH); 4.46 (d, J = 11.3, PhCH); 4.35 (br. d, J = 9.7, H–C(3)); 4.18 (d, J = 1.2, OH); 4.10 (m, H–C(4), H–C(5)); 3.86 (s, COOMe); 3.79 (dd, J = 10.5, 2.1, 1 H–C(9)); 3.77 (dd, J = 8.4, 1.3, H–C(7)); 3.71 (dt, J = 8.6, 2.6, H–C(8)); 3.66 (dd, J = 10.1, 2.8, 1 H–C(9)); 1.73 (s, AcN). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 169.82 (s); 168.56 (s); 138.12 (s); 137.99 (s); 137.86 (s); 137.77 (s); 128.7–127.6 (m, 20 d); 9.5.47 (s); 79.64 (d); 74.80 (d); 74.44 (d); 74.27 (t); 73.32 (t); 72.31 (t); 70.47 (d); 67.97 (t); 53.93 (q); 52.35 (d); 51.89 (d); 23.36 (d), CI–NIS: 682 (100, [M + 1 - Br]<sup>-</sup>). Anal. calc. for C<sub>40</sub>H<sub>44</sub>BrNO<sub>9</sub> (762.70): C 62.99, H 5.81, Br 10.48, N 1.84; found: C 63.09, H 5.83, Br 10.28, N 1.78.

Data of 13.  $R_t$  (AcOEt/hexane 1:1) 0.44.  $[\alpha]_{25}^{25} = +11.9$  (c = 0.9, CHCl<sub>3</sub>) ([10]:  $[\alpha]_D = 15.4$  (c = 1, CHCl<sub>3</sub>)). IR (CHCl<sub>3</sub>): 3500m, 3430m, 3000m, 2960m, 2880m, 1725s, 1680s, 1560m, 1550m, 1450m, 1370m, 1290m, 1240m, 1160s, 1130s, 1095s, 1050s, 1030s. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): 7.4–7.2 (m, 20 arom. H); 4.73 (d, J = 11.8, PhCH); 4.68 (d, J = 11.5, PhCH); 4.65–4.55 (m, 4 H, PhCH); 4.61 (d, J = 3.5, H–C(3)); 4.54 (d, J = 12.0, PhCH); 4.52 (dd, J = 10.9, 1.7, H–C(6)); 4.51 (d, J = 8.2, NH); 4.45 (dd, J = 10.1, 3.5, H–C(4)); 4.35 (d, J = 11.8, PhCH); 4.05–3.95 (m, 2 H, H–C(8), 1 H–C(9)); 3.83 (dt, J = 8.2, 10.3, H–C(5)); 3.80 (dd, J = 6.8, 1.7, H–C(7)); 3.78 (s, COOMe); 3.75 (dd, 11.6, 5.0, 1 H–C(9)); 3.51 (s, OH); 1.66 (s, AcN). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 170.77 (s); 168.20 (s); 138.67 (s); 138.07 (s); 137.71 (s); 129.1–127.5 (m, 20d); 96.16 (s); 78.38 (d); 24.25 (d); 73.36 (t); 72.64 (t); 72.26 (d); 70.60 (t); 70.26 (d); 69.64 (t); 53.16 (d); 52.76 (q); 49.46 (d); 23.43 (q). CL-MS: 682 (100, [M + 1 - Br]<sup>+</sup>). Anal. calc. for C<sub>40</sub>H<sub>44</sub>BrNO<sub>9</sub> (762.70) C 62.99, H 5.81, Br 10.48, N 1.84; found: C 62.84, H 5.56, Br 10.73, N 2.01.

*Methyl* 5-Acetamido-2,3-anhydro-4,7,8,9-tetra-O-benzyl-5-deoxy-β-D-erythro-L-gluco-2-nonulopyranosonate (14). DBU (0.77 ml, 5.17 mmol, 1.2 equiv.) was added to a soln. of 13 (3.30 g, 4.33 mmol) in abs. MeCN (25 ml). After stirring at r.t. for 10 min., the soln. was evaporated and the residue purified by chromatography (SiO<sub>2</sub>, AcOEt/hexane 1:1) to give 14 (2.507 g, 85%). Foam.  $R_{\rm f}$  (AcOEt/hexane 1:1) 0.32.  $[\alpha]_{\rm D}^{25}$ -19.6 (c = 1.1, CHCl<sub>3</sub>). IR (CHCl<sub>3</sub>): 3430m, 3000m, 2960m, 2870m, 1755s, 1680s, 1500m, 1450m, 1370m, 1310m, 1250m, 1170m, 1095s, 1070s, 1030m, 990w, 910w. 'H-NMR (400 MHz, CDCl<sub>3</sub>): 7.5–7.3 (m, 20 arom. H; 5.10 (d, J = 8.2, NH); 4.75 (d, J = 12.0, PhCH); 4.64 (d, J = 11.5, PhCH); 4.55–4.63 (m, 6 H, PhCH); 3.87 (d, J = 10.8, 3.4, 1 H–C(9)); 3.70 (dd, J = 10.5, 4.4, 1 H–C(9)); 3.70 (s, COOMe); 3.64 (s, H–C(3)); 1.74 (s, AcN). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 169.35 (s); 165.60 (s); 138.45 (s); 138.25 (s); 138.11 (s); 137.04 (s); 128.5–127.5 (m, 20 d); 79.51 (s); 77.88 (d); 75.98 (d); 73.90 (d); 73.52 (t); 73.32 (t); 72.12 (t); 71.96 (d); 68.85 (t); 57.35 (d); 52.89 (q); 47.96 (d); 22.33 (q). CI–MS: 682 (100, [M+1]<sup>+</sup>), 610 (15), 592 (25). Anal. calc. for C<sub>40</sub>H<sub>43</sub>NO<sub>9</sub> (681.79) C 70.47, H 6.36, N 2.05; found: C 70.32, H 6.51, N 1.88.

Methyl 5-Acetamido-2,7-anhydro-4,8,9-tri-O-benzyl-5-deoxy- $\beta$ -D-erythro-1-gluco-2-nonulopyranosonate (15) and Methyl 5-Acetamido-4,7,8,9-tetra-O-benzyl-2-bromo-2,5-dideoxy- $\beta$ -D-erythro-L-gluco-2-nonulopyranosonate (16). a) A soln. of freshly distilled TiBr<sub>4</sub> (1.78 g, 4.84 mmol, 1.1 equiv.) in abs. 1,2-

dichloroethane (20 ml) was added dropwise to a soln. of **14** (3.0 g, 4.4 mmol) in abs. 1,2-dichloroethane (50 ml) at -40°. After stirring at -40° for 5 min, the soln. was poured into a sat. Na<sub>2</sub>SO<sub>4</sub> soln. (50 ml), and AcOEt (150 ml) was added. The org. layer was treated with a 5% NaHCO<sub>3</sub> soln. (50 ml) and brine (20 ml), dried (MgSO<sub>4</sub>), and evaporated. Chromatography of the residue (SiO<sub>2</sub>, AcOEt/hexane 1:1, AcOEt after the elution of the first product) gave **16** (2.05 g, 61%) and **15** (0.91 g, 34%) as foams.

b) A soln. of freshly distilled TiBr<sub>4</sub> (1.426 g, 3.87 mmol, 1.1 equiv.) in abs. THF (30 ml) was added dropwise to a soln. of 14 (2.40 g, 3.52 mmol) in abs. THF (80 ml) at  $-70^{\circ}$ . After stirring at  $-70^{\circ}$  for 10 min, the soln. was directly poured on a silica-gel column. Elution with AcOEt/hexane 1:1 gave 16 (2.57 g, 96%) as a foam.

*Data of* **15**:  $R_t$  (AcOEt) 0.38.  $[\alpha]_D^{25} = -4.0$  (c = 1.1, CHCl<sub>3</sub>). IR (CHCl<sub>3</sub>): 3460 (br.), 3430*m*, 3000*m*, 2960*m*, 2860*m*, 1750*s*, 1670*s*, 1500*m*, 1450*m*, 1370*m*, 1310*w*, 1110*s*, 1090*s*, 1070*s*, 1030*s*, 950*w*, 910*w*. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): 7.35–7.25 (*m*, 15 arom. H); 6.37 (*d*, J = 9.1, NH); 4.75 (*d*, J = 11.8, 2 H, PhC*H*); 4.65–4.50 (*m*, 6 H, PhC*H*, H–C(6), H–C(7)); 4.28 (*dq*, J = 9.2, 1.6, H–C(5)); 4.06 (br. *d*, J = 4.4, H–C(3)); 3.81 (*s*, COOMe); 3.78 (*dd*, J = 10.4, 2.3, H–C(9)); 3.61 (*dd*, J = 10.4, 5.0, H–C(9)); 3.56 (*ddd*, J = 7.7, 5.0, 2.6, H–C(8)); 3.53 (*q*, J = 1.4, H–C(4)); 2.89 (*d*, J = 4.5, OH); 1.99 (*s*, AcN). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 169.43 (*s*); 167.55 (*s*); 138.16 (*s*); 138.03 (*s*); 137.47 (*s*); 128.83–127.52 (*m*, 15 *d*); 103.66 (*s*); 79.06 (*d*); 78.93 (*d*); 78.07 (*d*); 76.60 (*d*); 73.30 (*t*); 72.48 (*t*); 71.61 (*t*); 69.10 (*t*); 68.84 (*d*); 52.97 (*q*); 47.17 (*d*); 23.12 (*q*). CI-MS: 592 (100, [*M* + 1]<sup>+</sup>). Anal. calc. for  $C_{33}H_{37}NO_9$  (591.66): C 66.99, H 6.30, N 2.37; found: C 67.08, H 6.36, N 2.22.

*Data of* **16**:  $R_t$  (AcOEt/hexane 1:1) 0.39.  $[\alpha]_D^{25} = -53.6$  (c = 0.94, CHCl<sub>3</sub>). IR (CHCl<sub>3</sub>): 3550w, 3430m, 3000m, 2960m, 2860m, 1730s, 1680s, 1500m, 1440m, 1440m, 1370m, 1310m, 1280m, 1100s, 1040 (sh), 1025m, 910w, 890w. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): 7.40–7.25 (m, 20 arom. H); 4.95 (d, J = 11.9, PhCH); 4.7–4.6 (m, 5 H, NH, PhCH); 4.59 (d, J = 12.1, PhCH); 4.56 (d, J = 12.1, PhCH); 4.51 (d, J = 10.6, PhCH); 4.49 (br. d, J = 10.6, H–C(6)); 4.11 (dt, J = 10.7, 9.5, H–C(5)); 3.94 (dd, J = 9.7, 8.1, H–C(4)); 3.86 (s, COOMe); 3.82–3.74 (m, H–C(3), H–C(7), H–C(8), 1 H–C(9), OH); 3.67 (br. d, J = 11.0, 1 H–C(9)); 1.73 (s, AcN). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 169.90 (s); 167.64 (s); 138.31 (s); 138.21 (s); 137.85 (2 s); 127.50–128.87 (m, 20 d); 99.81 (s); 78.86 (d); 7.00 (d); 75.20 (d); 74.74 (d); 74.44 (d); 74.19 (t); 74.05 (t); 73.26 (t); 72.65 (t); 68.13 (t); 53.56 (q); 49.09 (d); 23.47 (q). CI-MS: 682 (17, [M + 1 - Br]<sup>+</sup>), 592(39), 576(23), 502(19), 486(27), 91(100). Anal. calc. for C<sub>40</sub>H<sub>44</sub>BrNO<sub>9</sub> (762.70) C 62.99, H 5.81, Br 10.48, N 1.84; found: C 62.81, H 5.68, Br 10.36, N 1.67.

*Methyl* 5-Acetamido-2,6-anhydro-4,7,8,9-tetra-O-benzyl-5-deoxy-D-arabino-L-gulo-2-nononate (17). A mixture of **16** (4.31 g, 5.65 mmol), 2,2'-dimethyl-2,2'-azobis[propanenitrile] (AIBN; 460 mg, 2.8 mmol, 0.5 equiv.), Bu<sub>3</sub>SnH (2.1 ml, 7.92 mmol, 1.4 equiv.) and dry benzene (150 ml) was heated to 60° for 30 min. Evaporation and chromatography of the residue (SiO<sub>2</sub>, AcOEt/hexane 1:4 to 2:1) gave **17** (3.529 g, 91%) as a foam.  $R_r$  (AcOEt/hexane 1:1) 0.25.  $[\alpha]_D^{25} = -9.5$  (c = 0.94, CHCl<sub>3</sub>). IR (CHCl<sub>3</sub>): 3550m (br.), 3430m, 3060w, 3000m, 2960m, 2870m, 1730s, 1675s, 1510m, 1500m, 1450m, 1440m, 1370m, 1320w, 1270s, 1200s (br.), 1100s, 1050s, 910w, 890w. 'H-NMR (400 MHz, CDCl<sub>3</sub>): 7.40–7.25 (m, 20 arom. H); 4.88 (d, J = 11.8, PhCH); 4.70 (d, J = 11.6, PhCH); 4.63–4.53 (m, 6 H, PhCH); 4.31 (d, J = 8.2, NH); 4.09 (dd, J = 10.4, 1.4, H–C(6)); 3.94 (dd, J = 9.8, 8.7, H–C(4)); 3.92–3.84 (m, H–C(8), 1 H–C(9)); 3.85 (dt, J = 2.1, 9.1, H–C(2)); 3.47 (dt, J = 8.2, 10.1, H–C(5)); 3.26 (d, J = 2.2, OH); 1.58 (s, AcN). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 170.32 (s); 169.98 (s); 138.11 (s); 138.08 (s); 129.35–127.53 (m, 20 d); 81.01 (d); 77.90 (d); 77.45 (d); 76.45 (d); 74.39 (t); 73.94 (d); 73.36 (t; 73.17 (t); 72.75 (d); 72.59 (t); 68.94 (t); 52.35 (q); 51.99 (d); 23.51 (q). CI-MS: 684 (100, [M + 1]<sup>+</sup>), 592 (20), 501 (16). Anal. calc. for C<sub>40</sub>H<sub>45</sub>NO<sub>9</sub> ·  $\frac{1}{2}H_2O$  (692.81): C 69.35, H 6.69, N 2.02; found: C 69.50, H 6.81, N 2.02

*Methyl* 5-Acetamido-2,6-anhydro-3,4,7,8,9-penta-O-benzyl-5-deoxy-D-arabino-L-gulo-2-nononate (18). A mixture of 17 (3.5 g, 5.12 mmol), NaH (300 mg, 12.5 mmol), and abs. DMF (130 ml) was stirred at r.t. for 1 h. Benzyl bromide (850 µl, 7.14 mmol) was then added and stirring continued for 16 h. After addition of *Dowex* 50 *WX4* (H<sup>+</sup> form), the partly hydrolyzed methyl ester was regenerated by treating the soln. with an Et<sub>2</sub>O soln. of diazomethane. The soln. was then poured into H<sub>2</sub>SO<sub>4</sub> (0.2*m*, 100 ml) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (4 × 100 ml). The org. layer was dried (MgSO<sub>4</sub>) and evaporated and the residue purified by chromatography (SiO<sub>2</sub>, AcOEt/hexane 1:1) to give 18 (2.5 g, 63%) and 17 (923 mg, 26%).  $R_1$  (AcOEt/hexane 1:1) 0.38.  $[\alpha]_D^{25} = -6.4$  (c = 1.31, CHCl<sub>3</sub>). IR (CHCl<sub>3</sub>): 3430m, 3090w, 3070w, 3000w, 2960m, 2870m, 1750s, 1680s, 1650m, 1500m, 1450m, 1440m, 1365m, 1250s (br.), 1095s, 1030s, 910m. 'H-NMR (400 MHz, CDCl<sub>3</sub>): 7.40–7.25 (m, 25 arom. H); 4.83 (d, J = 11.5, PhCH); 4.78 (d, J = 10.8, PhCH); 4.07 (d, J = 10.9, PhCH); 4.01 (d, J = 10.4, PhCH); 4.01 (d, J = 10.5, 1.4, PhCH); 4.01 (d, J = 10.5, 1.4, H–C(7)); 3.70 (s, COOMe); 3.69 (dd, J = 10.5, 3.5, H–C(9)); 1.60 (s, AcN). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 170.00 (s); 168.88 (s); 138.32 (s); 138.11 (s); 138.02 (s); 137.87 (s); 129.09–127.66 (m, 25 d; 82.23 (d); 80.05 (d); 78.11 (d);

77.45 (*d*); 76.51 (*d*); 74.73 (2 *t*); 74.21 (*d*); 73.39 (*t*); 73.33 (*t*); 72.60 (*t*); 68.54 (*t*); 52.15 (*q*); 51.95 (*d*); 23.50 (*q*). CI-MS: 774 (100,  $[M + 1]^+$ ). Anal calc. for C<sub>47</sub>H<sub>51</sub>NO<sub>9</sub> (773.93): C 72.94, H 6.64, N 1.81; found: C 72.94, H 6.45, N 1.55.

4-Acetamido-2,3,6,7,8-penta-O-benzyl-4-deoxy-β-D-erythro-L-gluco-octopyranosyl Acetate (19) and 4-Acetamido-2,3,6,7,8-penta-O-benzyl-4-deoxy-α-D-erythro-L-gluco-octopyranosyl Acetate (20). A soln. of 18 (1.55 g, 2.00 mmol) in MeOH (20 ml) and 1M aq. NaOH (3 ml, 3.00 mmol) was stirred at r.t for 2.5 h. Filtration of the soln. through a short *Dowex 50 WX4* (H<sup>+</sup> form) column (6 ml of resin) and evaporation of the solvents gave the free acid (1.49 g, 98%) which was used without further purification. A mixture of the free acid (1.60 g, 2.1 mmol), abs. toluene (15 ml), pyridine (1.5 ml), and Pb(OAc)<sub>4</sub> (2.80 g, 6.32 mmol, 3equiv.) was heated under N<sub>2</sub> at 60° for 2 h. Evaporation of the solvents at 0.01 Torr and chromatography of the residue (SiO<sub>2</sub>, AcOEt/hexane 1:1) gave **19/20**<sup>§</sup> (1.20 g, 74%). A sample of this mixture was separated by chromatography (SiO<sub>2</sub>, AcOEt/ hexane 1:2).

*Data of* **19**:  $R_t$  (AcOEt/hexane 1:1) 0.47.  $[\alpha]_D^{2s} = -27.4$  (c = 1.05, CHCl<sub>3</sub>). IR (CHCl<sub>3</sub>): 3430m, 3060w, 3000w, 2940m, 2870m, 1745s, 1680s, 1500m, 1450m, 1370m, 1270m, 1230s (br.), 1145m, 1095s, 1025s, 1010s, 930m. 'H-NMR (400 MHz, CDCl<sub>3</sub>): 7.37–7.27 (m, 25 arom. H); 6.33 (d, J = 3.5, H–C(1)); 4.89 (d, J = 11.7; PhCH); 4.70–4.50 (m, 8 H, NH, 7 PhCH); 4.52 (d, J = 10.8, PhCH); 4.46 (d, J = 11.7, PhCH); 4.34 (br. d, J = 10.7, H–C(5)); 4.04 (t, J = 9.6, H–C(3)); 3.90 (dt, J = 10.4, 9.6, H–C(4)); 3.80–3.70 (m, H–C(6), H–C(7), H–C(8)); 3.69 (dd, J = 9.2, 3.5, H–C(2)); 3.67 (m, H–C(8)); 1.89 (s, ACO); 1.72 (s, AcN). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 169.89 (s); 169.36 (s); 138.65 (s); 138.48 (s); 137.94 (2s); 137.61 (s); 129.50–127.23 (m, 25 d); 90.11 (d); 79.22 (d); 77.95 (d); 77.38 (d); 74.92 (t); 7.37 (t); 7.333 (t); 73.06 (t); 72.11 (t); 70.32 (d); 67.84 (t); 51.23 (d); 23.69 (q); 20.73 (q). CI-MS: 774 (5, [M + 1]<sup>+</sup>), 714 (100), 624 (30). Anal. calc. for C<sub>47</sub>H<sub>51</sub>NO<sub>9</sub> (773.93): C 72.94, H 6.64, N 1.81; found: C 72.91, H 6.47, N 1.75.

*Data of* **20**:  $R_{\rm t}$  (AcOEt/hexane 1:1) 0.51.  $[\alpha]_{25}^{25} = -3.4$  (c = 1.1, CHCl<sub>3</sub>). IR (CHCl<sub>3</sub>): 3430m, 3090w, 3030w, 3000w, 2920m, 2870m, 1755s, 1680s, 1510m, 1500m, 1450m, 1370m, 1325w, 1220m, 1090s, 1050s, 1030s, 910w, 890w. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): 7.4–7.2 (m, 25 arom. H); 5.57 (d, J = 8.2, H–C(1)); 4.39 (d, J = 8.0, NH); 4.85–4.50 (m, 10 H, PhCH); 4.06 (br. d, J = 9.7, H–C(5)); 3.90–3.84 (m, H–C(3), H–C(4)); 3.80 (ddd, J = 8.9, 3.3, 2.2, H–C(7)); 3.75 (dd, J = 10.7, 2.2, 1 H–C(8)); 3.70 (dd, J = 8.9, 1.0, H–C(6)); 3.67 (dd, J = 10.7, 3.3, 1 H–C(8)); 3.62 (t, J = 8.5, H–C(2)); 2.03 (s, AcO); 1.55 (s, AcN). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 169.77 (s); 169.01 (s); 138.34 (s); 138.22 (s); 138.00 (s); 137.96 (s); 137.93 (s); 129.39–127.51(m, 25 d); 94.09 (d); 81.38 (d); 80.98 (d); 77.34 (d); 74.92 (t); 74.82 (t); 73.72 (t); 73.65 (d); 73.23 (t); 72.82 (t); 72.58 (d); 68.02 (t); 51.35 (d); 23.51 (q); 20.89 (q). CI-MS: 774 (8, [M + 1]\*), 714 (100), 624 (15), 594 (9), 534 (8). Anal. calc. for C<sub>47</sub>H<sub>51</sub>NO<sub>9</sub> (773.93): C 72.94, H 6.64, N 1.81; found: C 72.73, H 6.65, N 1.90.

4-Acetamido-2,3,6,7,8-penta-O-benzyl-4-deoxy-D-erythro-L-gluco-octopyranose (21). A soln. of 19/20 (1,10 g, 1.42 mmol), abs. MeOH (40 ml), and 0.5M NaOMe in MeOH (1 ml; 0.5 mmol) was stirred at r.t. for 2.5 h. Filtration through a *Dowex 50 WX4* (H<sup>+</sup> form) column and evaporation gave 21 (100%).  $\alpha$ -D/ $\beta$ -D 1:1.9,  $R_t$  (AcOEt/hexane 1:1) 0.32. 'H-NMR (400 MHz, CDCl<sub>3</sub>): 7.4–7.2 (m, 25 arom. H); 5.06 (t, J = 2.9, 0.65 H, H–C(1)); 4.9–4.4 (m, 10 H, PhCH); 4.74 (d, J = 9.6, 0.65 H, NH); 4.41 (d, J = 8.5, 0.35 H, NH); 4.35 (dd, J = 7.3, 6.0, 0.35 H, H–C(1)); 4.18 (br. d, J = 10.7, 0.65 H, H–C(5)); 4.09 (q, J = 9.8, 0.65 H, H–C(4)); 3.84 (0.35 H, H–C(5)); 3.83 (t, J = 9.1, 0.65 H, H–C(3)); 3.80 (t, J = 8.8, 0.35 H, H–C(2)); 3.29 (dd, J = 7.8, 0.65 H, H–C(6)); 3.68 (q, J = 9.4, 0.35 H, H–C(4)); 3.58 (dd, J = 9.1, 3.5, 0.65 H, H–C(2)); 3.29 (dd, J = 2.7, 0.65 H, H–C(2)); 3.64–3.73, 3.77–3.93 (2m, all missing signals); 2.58 (d, J = 6.0, 0.35 H, OH); 2.42 (d, J = 2.7, 0.65 H, OH); 1.74 (s, 1.95 H, AcN); 1.62 (s, 1.05 H, AcN). CI-MS: 732 (4z, [M + 1]<sup>+</sup>), 714 (56), 624 (29), 516 (18), 91 (100).

O-[4-Acetamido-2,3,6,7,8-penta-O-benzyl-4-deoxy- $\alpha$ -D-erythro-L-gluco-octopyranosyl] Trichloroacetimidate (22) and O-[4-Acetamido-2,3,6,7,8-penta-O-benzyl-4-deoxy- $\beta$ -D-erythro-L-gluco-octopyranosyl] Trichloroacetimidate (23). A mixture of 21 (500 mg, 0.683 mmol), K<sub>2</sub>CO<sub>3</sub> (450 mg, 3.261 mmol, 4.8 equiv.), trichloroacetonitrile (344 µl, 3.414 mmol, 5 equiv.), and abs. CH<sub>2</sub>Cl<sub>2</sub> (20 ml) was stirred at r.t. After 48 h, a second portion of trichloroacetonitrile (344 µl) was added. After stirring for another 48 h, the soln. was poured onto a silica-gel column. Elution (AcOEt/hexane 1:2) gave 22 (295 mg, 49.3%), 23 (123 mg, 20.5%), and 21 (70 mg, 14 %).

*Data of* **22**:  $R_{\rm f}$  (AcOEt/hexane 1:1) 0.38.  $[\alpha]_{\rm D}^{25} = -5.4$  (c = 0.8, CHCl<sub>3</sub>). IR (CHCl<sub>3</sub>): 3430*m*, 3340*m*, 3090*w*, 3060*w*, 3000*m*, 2920*m*, 2860*m*, 1670*s*, 1090*s*, 1030*m*, 830*m*. 'H-NMR (400 MHz, CDCl<sub>3</sub>): 8.63 (*s*, NH); 7.4–7.2 (*m*, 25 arom. H); 5.78 (*d*, J = 8.3, H–C(1)); 4.97 (*d*, J = 10.8, PhCH); 4.83 (*d*, J = 11.7, PhCH); 4.76 (*d*, J = 11.0, PhCH); 4.64–4.64 (*m*, 7 H, PhCH); 4.38 (*d*, J = 8.7, NH); 4.15 (*dd*, J = 10.5, 1.3, H–C(5)); 4.00 (*t*, J = 9.4,

<sup>&</sup>lt;sup>8</sup>) The 19/20 ratio was determined by integration of the H–C(1) signals of 19 and 20 in the <sup>1</sup>H-NMR spectra of the crude product.

H–C(3)); 3.89 (*ddd*, *J* = 7.8, 3.8, 2.0, H–C(7)); 3.83 (*dd*, *J* = 10.8, 2.3, 1 H–C(8)); 3.80–3.67 (*m*, H–C(2), H–C(4), H–C(6), 1 H–C(8)); 1.56 (*s*, AcN). <sup>13</sup>C-NMR (50 MHz, CDCI<sub>3</sub>): 169.88 (*s*); 161.35 (*s*); 138.58 (*s*); 138.44 (*s*); 138.18 (*2s*); 137.98 (*s*); 129.33–127.58 (*m*, 25*d*); 98.61 (*d*); 90.90 (*s*); 81.45 (*d*); 80.48 (*d*); 78.21 (*d*); 74.97 (*t*); 74.88 (*t*); 74.07 (*d*); 73.35 (*2t*); 73.01 (*d*); 72.89 (*t*); 68.89 (*t*); 51.99 (*d*); 23.56 (*q*). CI-MS: 714 (42), 624 (50), 606 (37), 91 (100). Anal. calc. for  $C_{47}H_{49}Cl_{3}N_2O_8$  (876.28): C 64.42, H 5.64, CI 12.14, N 3.20; found: C 64.60, H 5.37, CI 12.29, N 3.40.

*Data of* **23**:  $R_t$  (AcOEt/hexane 1:1) 0.41.  $[\alpha]_D^{2s} = -21.8$  (c = 0.97, CHCl<sub>3</sub>). IR (CHCl<sub>3</sub>): 3430m, 3350m, 3090w, 3060w, 3000m, 2930m, 2870m, 1670s, 1510m, 1500m, 1455m, 1370m, 1295m, 1115s, 1090s, 1070s, 1030s, 970m, 910w, 895w, 880w, 830w. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): 8.53 (s, NH); 7.4–7.2 (m, 25 arom. H); 6.51 (d, J = 3.2, H–C(1)); 4.87 (d, J = 11.7, PhCH); 4.73–4.45 (m, 10 H, NH, PhCH); 4.27 (br. d, J = 10.2, H–C(5)); 4.11 (q, J = 9.8, H–C(4)); 4.06 (t, J = 9.8, H–C(3)); 3.79 (dd, J = 9.0, 3.2, H–C(2)); 3.65 (br. s, H–C(6), H–C(7)); 3.65 (br. s, 2 H–C(8)); 1.76 (s, AcN). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 169.85 (s); 160.97 (s); 138.35 (s); 138.26 (s); 137.89 (2s); 137.80 (s); 129.00–127.49 (m, 25 d); 94.60 (d); 91.15 (s); 79.64 (d); 77.42 (d); 76.89 (d); 74.52 (t); 74.40 (d); 74.14 (t); 73.14 (t); 72.69 (t); 72.16 (t); 71.43 (d); 68.26 (t); 50.43 (d); 23.58 (q). CI-MS: 714 (5), 624 (43), 534 (28), 516 (31), 444 (10), 426 (21) 107 (100), 91 (74). Anal. calc. for C<sub>47</sub>H<sub>49</sub>Cl<sub>3</sub>N<sub>2</sub>O<sub>8</sub> (876.28): C 64.42, H 5.64, CI 12.14, N 3.20; found: C 64.27, H 5.84, CI 12.35, N 3.12.

Dimethyl (4-Acetamido-2,3,6,7,8-penta-O-benzyl-4-deoxy- $\alpha$ -D-erythro-L-gluco-octopyranosyl)phosphonate (24), Dimethyl (4-Acetamido-2,3,6,7,8-penta-O-benzyl-4-deoxy- $\beta$ -D-erythro-L-glucooctopyranosyl)phosphonate (25), and 4-Acetamido-1,6-anhydro-2,3,7,8-tetra-O-benzyl-4-deoxy- $\alpha$ -D-erythro-Lgluco-octopyranose (26). A mixture of 22 (277 mg, 0.316 mmol), P(MeO)<sub>3</sub> (250 µl, 2.12 mmol, 6.7 equiv.), molecular sieves (4Å, 2.8 g) and abs. CH<sub>2</sub>Cl<sub>2</sub> (5 ml) was stirred at r.t. Me<sub>3</sub>SiOTf (58 µl, 0.32 mmol, 1.01 equiv.) was added at 0° and stirring was continued for 24 h at 5°. The soln. was filtered through Celite and the filtrate evaporated. Chromatography of the residue (SiO<sub>2</sub>, AcOEt/hexane 1:1) gave 24/25 (113 mg, 44%) and 26 (80 mg, 41%). Enriched fractions of 24 and 25 were collected and their <sup>1</sup>H-NMR spectra measured.

*Data of* **24:**  $R_i$  (AcOEt/hexane 3:1) 0.39. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): 7.4–7.2 (*m*, 25 arom. H); 4.90 (*d*, J = 10.3, PhCH); 4.84 (*d*, J = 11.6, PhCH); 4.83 (*d*, J = 10.3, PhCH); 4.68 (*d*, J = 11.2, PhCH); 4.62–4.50 (*m*, 7 H, PhCH, NH); 3.9–3.8 (*m*, 6 H); 3.76 (*d*, J(H,P) = 10.6, 3H, P(OMe)\_2); 3.72–3.63 (*m*, 3 H); 3.68 (*d*, J(H,P) = 10.7, 3 H, P(OMe)\_2); 1.62 (*s*, AcN). <sup>31</sup>P-NMR (81 MHz, CDCl\_3): 22.60.

*Data of* **25**:  $R_1$  (AcOEt/hexane 3:1) 0.36. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): 7.4–7.2 (*m*, 25 arom. H); 5.85 (*d*, *J* = 9.6, NH); 4.76 (*d*, *J* = 11.7, PhCH); 4.71 (*d*, *J* = 11.3, PhCH); 4.71 (*d*, *J* = 11.3, PhCH); 4.68 (*d*, *J* = 11.6, PhCH); 4.66 (*d*, *J* = 11.2, PhCH); 4.62 (*d*, *J* = 10.0, PhCH); 4.59 (*d*, *J* = 11.2, PhCH); 4.58 (*d*, *J* = 10.0, PhCH); 4.55 (*d*, *J* = 12.0, PhCH); 4.51 (*dd*, *J*(H,P) = 12.0, *J* = 5.5, H–C(1)); 4.49 (*d*, *J* = 12.1, PhCH); 4.34 (*q*, *J* = 8.5, H–C(4)); 4.17 (*ddd*, *J* = 8.4, 3.1, 1.2, H–C(5)); 4.06 (*dt*, *J* = 1.1, 7.2, H–C(3)); 3.93 (*ddd*, *J*(H,P) = 23.1, *J* = 7.4, 5.5, H–C(2)); 3.90–3.84 (*m*, H–C(6), H–C(7), 1 H–C(8)); 3.71 (*dd*, *J* = 10.6, 5.0, 1 H–C(8)); 3.67 (*d*, *J*(H,P) = 10.8, 3 H, P(OMe)\_2); 1.36 (*d*, *J*(H,P) = 10.7, 3 H, P(OMe)\_2); 1.76 (*s*, AcN). <sup>31</sup>P-NMR (81 MHz, CDCl<sub>3</sub>): 23.99.

*Data of* **26**:  $R_t$  (AcOEt/hexane 1:1) 0.36. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): 7.4–7.2 (*m*, 20 arom. H); 6.25 (*d*, J = 9.4, NH); 5.40 (br. *s*, H–C(1)); 4.75 (*d*, J = 11.7, PhCH); 4.73 (*d*, J = 12.3, PhCH); 4.60–4.50 (*m*, 4 H, PhCH); 4.51 (br. *d*, J = 7.7, H–C(6)); 4.36 (br. *s*, H–C(5)); 4.33 (*d*, J = 11.9, PhCH); 4.30 (*d*, J = 11.9, PhCH); 4.23 (*d*, fine struct., J = 9.1, H–C(4)); 3.76 (*dd*, J = 10.4, 2.7, 1 H–C(8)); 3.60 (*dd*, J = 10.5, 5.3, 1 H–C(8)); 3.46 (*ddd*, J = 7.7, 5.3, 2.7, H–C(7)); 3.44 (br. *s*, H–C(2) or H–C(3)); 3.32 (br. *s*, H–C(2) or H–C(3)); 1.93 (*s*, AcN). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 169.06 (*s*); 138.23 (*s*); 138.18 (*s*); 137.63 (*s*); 137.25 (*s*); 128.51–127.50 (*m*, 20 *d*); 100.84 (*d*); 78.30 (*s*); 76.29 (*s*); 75.27 (*s*); 74.81 (*d*); 73.42 (*t*); 72.55 (*t*); 71.77 (*t*); 71.41 (*t*); 69.78 (*t*); 47.50 (*d*); 23.22 (*q*). CI-MS: 624 (100, [M + 1]<sup>+</sup>).

Dimethyl (4-Acetamido-2,3,6,7,8-penta-O-acetyl-4-deoxy- $\alpha$ -D-erythro-L-gluco-octopyranosyl)phosphonate (27) and Dimethyl (4-Acetamido-2,3,6,7,8-penta-O-acetyl-4-deoxy- $\beta$ -D-erythro-L-glucooctopyranosyl)phosphonate (28). A mixture 24/25 (100 mg, 0.121 mmol), 10% Pd/C (50 mg), and MeOH (15 ml) was hydrogenated at r.t./8 atm for 19 h. The soln. was then filtered through Celite and evaporated. TLC: single spot. <sup>1</sup>H-NMR: no arom. signals. The residue was acetylated overnight in Ac<sub>2</sub>O/pyridine 1:2 (3 ml) to give, after evaporation and chromatography (SiO<sub>2</sub>, AcOEt/MeOH 95:5), 27 (20 mg, 28%), 28 (22 mg, 31%), and 27/28 (27.0 mg, 38%).

Data of **27**:  $R_f$  (AcOEt/MeOH 9:1) 0.36.  $[\alpha]_{25}^{25} = -36.8$  (c = 0.75, CHCl<sub>3</sub>). IR (CHCl<sub>3</sub>): 3430*m*, 3000*m*, 2960*m*, 2860*w*, 1740*s*, 1685*s*, 1655 (sh), 1510*w*, 1370*s*, 1290*s*, 1240*s* (br.), 1080 (sh), 1040*s*, 940*w*, 905*w*, 835*m*. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): 5.36 (*dt*, *J*(H,P) = 9.2, *J* = 10.5, H–C(2)); 5.35 (*d*, *J* = 9.6, NH); 5.32 (*ddd*, *J* = 6.5, 2.1, 0.7, H–C(6)); 5.21 (*dt*, *J* = 2.6, 6.4, H–C(7)); 5.15 (*dt*, *J* = 0.7, 10.2, H–C(3)); 4.42 (*dd*, *J* = 12.5, 2.6, 1 H–C(8)); 4.13 (*q*, *J* = 10.3, H–C(4)); 4.11 (*dd*, *J* = 12.5, 6.3, 1 H–C(8)); 3.85 (*d*, *J*(H,P) = 10.7, P(OMe)<sub>2</sub>); 3.79 (*d*, *J*(H,P) = 10.9, 3 H, P(OMe)<sub>2</sub>); 3.76 (*dd*, *J* = 10.3, 2.1, H–C(5)); 3.75 (*t*, *J*(H,P) = 10.4, *J* = 10.4, H–C(1)); 2.13, 2.08, 2.05, 2.04,

2.03 (5 s, 5 AcO); 1.90 (s, AcN). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 171.07 (s); 170.54 (s); 170.28 (s); 170.22 (s); 170.12 (s); 169.26 (s); 78.38 (dd, J(C,P) = 16.2); 74.12 (dd, J(C,P) = 18.6); 72.82 (dd, J(C,P) = 173.2); 70.70 (d); 67.63 (d); 67.44 (d); 62.13 (t); 54.20 (dq, J(C,P) = 6.9); 53.15 (dq, J(C,P) = 6.6); 49.18 (d); 22.95 (q); 20.78 (q); 20.67 (2q); 20.62 (q); 20.60 (q). <sup>31</sup>P-NMR (81 MHz, CDCl<sub>3</sub>): 18.60. CI-MS: 584 (100, [M + 1]<sup>+</sup>).

Data of **28**:  $R_t$  (AcOEt/MeOH 9:1) 0.40.  $[\alpha]_D^{25} = +9.7$  (c = 0.82, CHCl<sub>3</sub>). IR (CHCl<sub>3</sub>): 3430*m*, 3000*m*, 2960*m*, 2860*m*, 1745*s*, 1690*s*, 1510*m*, 1370*s*, 1290*m*, 1240*s* (br.), 1080 (sh), 1035*s*, 950*w*, 905*w*, 860*m*, 830*m*. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): 5.67 (t, J = 10.0, H–C(3)); 5.64 (d; J = 10.3; NH); 5.33 (dd, J = 7.5, 2.1, H–C(6)); 5.16 (ddd, J(H,P) = 30.8, J = 9.9, 7.3, H–C(2)); 4.74 (dd, J(H,P) = 10.2, J = 7.4, H–C(1)); 4.38 (dd, J = 12.6, 2.9, 1 H–C(8)); 4.36 (dt, J = 10.3, 1.7, H–C(5)); 4.22 (q, J = 10.2, H–C(4)); 4.11 (dd, J = 12.4, 5.2, 1 H–C(8)); 3.78 (d, J(H,P) = 10.7, 3 H, P(OMe)<sub>2</sub>); 3.77 (d, J(H,P) = 10.9, 3 H, P(OMe)<sub>2</sub>); 2.13, 2.11, 2.09, 2.05, 2.04 (5 *s*, 5 AcO); 1.90 (*s*, AcN). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 170.80 (*s*); 170.53 (*s*); 170.19 (*s*); 170.08 (*s*); 169.83 (*s*); 169.70 (*s*); 74.02 (*d*); 70.60 (*d*); 69.86 (*d*); 69.07 (*d*); 67.91 (*d*, J(C,P) = 152.4); 67.28 (*d*); 61.76 (*t*); 53.74 (*dq*, J(C,P) = 7.4); 52.75 (*dq*, J(C,P) = 6.9); 48.69 (*d*); 23.00 (*q*); 21.03 (*q*); 20.78 (*q*); 20.67 (3 *q*). <sup>31</sup>P-NMR (81 MHz, CDCl<sub>3</sub>): 21.56. CI-MS: 584 (100, [M + 1]\*).

Diethyl (2,3,4,6-Tetra-O-acetyl- $\beta$ -D-glucopyranosyl)phosphonate (**30**) and Diethyl (2,3,4,6-Tetra-O-acetyl- $\alpha$ -D-glucopyranosyl)phosphonate (**31**). A mixture of crude **29** [15] (2.0 g, 3.03 mmol), 10% Pd/C (200 mg), and MeOH (50 ml) was hydrogenated at r.t./8 atm for 16 h. The soln. was then filtered through *Celite* and evaporated. The residue was acetylated overnight in Ac<sub>2</sub>O/Pyridine 1:2 (10 ml) to give, after evaporation and chromatography (SiO<sub>2</sub>, AcOEt), **30** (58 mg, 4%) and **31** (1.304 g, 92%) as colorless oils.

*Data of* **30**:  $R_t$  (AcOEt) 0.30. 'H-NMR (400 MHz, CDCl<sub>3</sub>): 5.34 (*dt*, *J*(H,P) = 10.3, *J* = 9.1, 10.3, H–C(2)); 5.20 (*dt*, *J*(H,P) = 0.6, *J* = 9.3, H–C(3)); 5.08 (*t*, *J* = 9.7, H–C(4)); 4.30–4.10 (*m*, 6 H, 2 H–C(6), P(OCH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>); 3.83 (*t*, *J*(H,P) = 10.2, *J* = 10.2, H–C(1)); 3.67 (*ddd*, *J* = 9.9, 4.9, 2.3, H–C(5)); 2.06, 2.04, 2.03, 2.01 (4 *s*, 3 H, AcO); 1.34 (*t*, *J*(H,P) = 7.1, 3 H, P(OCH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>); 1.34 (*t*, *J*(H,P) = 7.1, 3 H, P(OCH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 170.40 (*s*); 170.15 (*s*); 169.28 (*s*); 169.03 (*s*); 77.44 (*dd*, *J*(C,P) = 17.2); 74.14 (*dd*, *J*(C,P) = 17.9); 73.15 (*dd*, *J*(C,P) = 172.7); 67.90 (2 *d*); 63.42 (*dt*, *J*(C,P) = 7.3); 63.28 (*dt*, *J*(C,P) = 7.0); 61.97 (*t*); 20.61 (*q*); 20.56 (*q*); 20.50 (*q*); 20.46 (*q*); 16.35 (*2dq*, *J*(C,P) = 5.6). <sup>31</sup>P-NMR (81 MHz, CDCl<sub>3</sub>): 16.18.

Data of **31**:  $R_1$  (AcOEt) 0.39. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): 5.81 (t, J = 9.4, H–C(3)); 5.14 (ddd, J(H,P) = 29.6, J = 9.8, 7.4, H–C(2)); 5.02 (t, J = 9.4, H–C(4)); 4.66 (dd, J(H,P) = 11.4, J = 4.1, H–C(1)); 4.30–4.10 (m, H–C(5), 2 H–C(6), P(OCH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>); 2.09, 2.08, 2.03, 2.03 (4s, 4 AcO); 1.41 (t, J(H,P) = 7.1, 3 H, P(OCH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>); 1.36 (t, J(H,P) = 7.0, 3 H, P(OCH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 170.47 (s); 169.96 (s); 169.77 (s); 169.55 (s); 72.27 (d); 70.18 (d); 69.45 (dd, J(C,P) = 154); 69.04 (d); 68.16 (d); 63.45 (dt, J(C,P) = 7.0); 62.45 (dt, J(C,P) = 6.9); 62.07 (t); 20.58 (3q); 20.51 (q); 16.41 (dq, J(C,P) = 5.2); 16.31 (dq, J(C,P) = 4.9). <sup>31</sup>P-NMR (81 MHz, CDCl<sub>3</sub>): 19.74. CI-MS: 469 (100, [M + 1]\*), 427 (41), 385 (11), 157 (49).

Diethyl (2,3,4,6-Tetra-O-acetyl-1-C-bromo- $\alpha$ -D-glucopyranosyl)phosphonate (32). a) From 30: A mixture of 30 (100 mg, 0.213 mmol), NBS (90 mg, 0.506 mmol, 2.4 equiv.), K<sub>2</sub>CO<sub>3</sub>(60 mg, 0.434 mmol, 2.0 equiv.), and CCl<sub>4</sub> (7 ml) was heated to reflux and irradiated with a 100-W lamp for 2 h. After cooling, the soln. was filtered and evaporated and the residue purified by chromatography (SiO<sub>2</sub>, AcOEt): 105 mg of crude product. CCl<sub>4</sub> (1.5 ml) was added, the suspension cooled to 0°, and the precipitated succinimide (30 mg) was filtered off. Evaporation of the solvent gave 32 (75 mg, 64%).

b) *From* **31**: As described under *a*), a mixture of **31** (55 mg, 0.117 mmol), NBS (50 mg, 0.28 mmol, 2.4 equiv.),  $K_2CO_3(33 \text{ mg}, 0.24 \text{ mmol}, 2.0 \text{ equiv.})$ , and  $CCl_4$  (4 ml) gave **32** (34 mg, 53%).  $R_t$  (AcOEt) 0.45. <sup>1</sup>H-NMR (400 MHz, CDCl\_3): 5.47 (*t*, *J* = 9.5, H–C(3)); 5.38 (*dd*, *J*(H,P) = 6.2, *J* = 9.5, H–C(2)); 5.21 (*t*, *J* = 9.7, H–C(4)); 4.40–4.20 (*m*, H–C(5), H–C(6), P(OCH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>); 4.14 (*dd*, *J* = 12.4, 1.7, H–C(6)); 2.06, 2.05, 2.03, 1.98 (4 *s*, 4 AcO); 1.36 (*t*, *J*(H,P) = 6.9, 3 H, P(OCH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>); 1.35 (*t*, *J*(H,P) = 7.1, 3 H, P(OCH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 170.20 (*s*); 169.70 (*s*); 169.18 (*s*); 168.68 (*s*); 97.58 (*d*, *J*(C,P) = 195.6); 73.60 (*dd*, *J*(C,P) = 12.5); 71.49 (*dd*, *J*(C,P) = 12.3); 68.83 (*dd*, *J*(C,P) = 2.9); 66.60 (*d*); 65.71 (*dt*, *J*(C,P) = 5.8). <sup>3</sup>P-NMR (81 MHz, CDCl<sub>3</sub>): 80.6 CI-MS: 549 (100, [*M* + 1]<sup>+</sup>), 547 (100, [*M* + 1]<sup>+</sup>), 507 (22), 505 (20), 467 (20), 409 (15), 365 (45), 263 (28).

Diethyl (3,4,6-Tri-O-acetyl-2-deoxy-D-arabino-hex-1-enopyranosyl)phosphonate (33). a) From 32: A mixture of 32 (25 mg, 46  $\mu$ mol), EtOH (0.5 ml), and a Zn/Cu slurry (0.1 ml) was stirred at r.t for 30 min. The soln. was filtered and evaporated. Pyridine (2 ml) and Ac<sub>2</sub>O (1 ml) were added, and the soln. was stirred at r.t. for 2 h. Evaporation and chromatography (SiO<sub>2</sub>, AcOEt) afforded 33 (14.2 mg, 76%).  $R_f$  (AcOEt) 0.38. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): 5.80 (dd, J(H,P) = 10.5, J = 3.3, H–C(2)); 5.43 (dddd, J(H,P) = 2.2, J = 5.9, 3.3, 0.6, H–C(3)); 5.25 (dd, J = 7.5, 6.2, H–C(4)); 4.40 (ddd, J = 11.8, 5.2, 0.5, H–C(6)); 4.35 (dddd, J = 7.8, 5.4, 2.7, 0.5, H–C(5)); 4.23 (dd, J = 11.8, 2.8, H–C(6)); 4.40–4.20 (m, 4 H, P(OCH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>); 2.06, 2.05, 2.05 (3 s, 3 AcO); 1.35 (t,

 $J(H,P) = 7.1, 3 H, P(OCH_2CH_3)_2$ ; 1.34 ( $t, J(H,P) = 7.0, 3 H, P(OCH_2CH_3)_2$ ). <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>): 169.90 (s); 169.56 (s); 168.96 (s); 147.28 (d, J(C,P) = 225.2, C(1)); 110.78 (dd, J(C,P) = 22.7, C(2)); 74.66 (dd, J(C,P) = 9.1, C(5)); 66.37 (dd, J(C,P) = 14.6, C(3)); 66.02 (d, C(4)); 62.83 (m, 2 C, 2 POC); 60.47 (t, C(6)); 20.28 (q); 20.18 (q); 20.09 (q); 15.75 (2dq, J(C,P) = 5.8). <sup>31</sup>P-NMR (81 MHz, CDCl<sub>3</sub>): 6.70.

b) From **31**: A soln. of **31** (100 mg, 0.213 mmol) was heated to reflux under the following conditions: *i*) with 7-methyl-1,5,7-triazabicyclo[4.4.0]dec-5-ene (MTBD; 51.3  $\mu$ l, 0.416 mmol, 2 equiv. in toluene (3 ml) for 7h, *ii*) with 2-[(*tert*-butyl)imino]-2-(diethylamino)-perhydro-1,3-dimethyl-1,3,2-diazaphosphorine [26] (BEMP; 123  $\mu$ l, 0.416 mmol, 2 equiv.) in MeCN (3 ml) for 8 h; *iii*) with Bu<sub>4</sub>NF · 3H<sub>2</sub>O (202 mg, 0.64 mmol, 3 equiv.) in THF (3 ml) for 2 h. Evaporation and chromatography (SiO<sub>2</sub>, AcOEt) gave unseparable **31/33** mixtures (*i*)72 mg, 73% (9:1); *ii*) 34 mg, 36% (58:42); *iii*) 22 mg, 22% (95:5)).

*Dimethyl* (4-Acetamido-2,3,6,7,8-penta-O-acetyl-1-bromo-4-deoxy-D-erythro-β-L-gluco-octopyranosyl)phosphonate (**34**). A mixture of **27/28** (60 mg, 0.103 mmol), NBS (55 mg, 0.310 mmol, 3 equiv.), K<sub>2</sub>CO<sub>3</sub> (28 mg, 2 equiv.), and abs. CCl<sub>4</sub> (3.5 ml) was irradiated with a 100-W lamp under reflux for 3 h. Evaporation and chromatography (SiO<sub>2</sub>, AcOEt/MeOH 95:5 to 90:10) gave **34** (31 mg, 45%) and starting material (9 mg, 15%). *R*<sub>t</sub> (AcOEt/MeOH 9:1) 0.48. [ $\alpha$ ]<sub>D</sub><sup>25</sup> = -50.1 (*c* = 0.79, CHCl<sub>3</sub>). IR (CHCl<sub>3</sub>): 3430w, 3000w, 2960w, 2860w, 1750s, 1690s, 1505m, 1430w, 1370s, 1200–1250s (br.), 1140w, 990s, 950m, 890w, 865w, 840w. 'H-NMR (400 MHz, CDCl<sub>3</sub>): 5.44 (dd, J(H,P) = 6.5, 9.5, H–C(2)); 5.42 (ddd, *J* = 8.5, 2.1, 1.2, H–C(6)); 5.41 (d, *J* = 10.2, NH); 5.34 (ddd, *J* = 10.4, 9.4, 0.9, H–C(3)); 5.14 (ddd, *J* = 8.5, 5.1, 2.7, H–C(7)); 4.39 (q, *J* = 10.5, H–C(4)); 4.33 (ddd, *J* = 11.0, 2.1, 1.6, H–C(5)); 4.31 (dd, *J* = 12.7, 2.8, H–C(8)); 4.04 (dd, *J* = 12.6, 5.1, H–C(8)); 3.97 (d, *J*(H,P) = 10.6, 3 H, P(OMe)<sub>2</sub>); 3.91 (d, *J*(H,P) = 10.8, 3 H, P(OMe)<sub>2</sub>); 2.14, 2.12, 2.10, 2.05, 2.02 (5 s, 5 AcO); 1.92 (s, AcN). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 170.97 (s); 170.55 (s); 170.17 (s); 179.76 (s); 169.31 (s); 168.90 (s); 96.94 (d, *J*(C,P) = 195.5); 74.71 (dd, *J*(C,P) = 11.3); 71.72 (dd, *J*(C,P) = 12.9); 68.93 (d); 68.74 (dd, *J*(C,P) = 3.3); 66.22 (d); 61.72 (t); 56.32 (dq, *J*(C,P) = 7.0); 55.38 (dq, *J*(C,P) = 7.5); 47.86 (d); 22.94 (q); 21.04 (q); 20.67 (2q); 20.59 (q); 20.50 (q). <sup>31</sup>P-NMR (81 MHz, CDCl<sub>3</sub>): 9.46. CI-MS: 664 (14, [*M* + 1]<sup>+</sup>), 662 (14, [*M* + 1]<sup>+</sup>), 464 (100).

Dimethyl (4-Acetamido-3,6,7,8-tetra-O-acetyl-2,4-dideoxy-D-glycero-D-galacto-oct-1-enopyranosyl)phosphonate (**35**). To a soln. of **34** (29 mg, 0.044 mmol) and EtOH (99.5%, 1 ml), activated Zn/Cu slurry (200 µl; see preparation of **33**) was added under vigourous stirring. After 30 min, the Zn/Cu was filtered off and the filtrate evaporated. The residue was then dissolved in Ac<sub>2</sub>O (250 µl) and pyridine (500 µl) and stirred for 3 h at r.t. Evaporation and chromatography (SiO<sub>2</sub>, AcOEt/MeOH 92.5:7.5) gave **35** (16.5 mg, 72%).  $R_f$  (AcOEt/MeOH 9:1) 0.38.  $[\alpha]_p^{35} = +36.7$  (c = 0.83, CHCl<sub>3</sub>). IR (CHCl<sub>3</sub>): 3430w, 3040w, 3000m, 2960m, 2860w, 1745s, 1690s, 1650w, 1540 (sh), 1505m, 1430w, 1370s, 1200–1250s (br.), 1145s, 1100s, 1040s, 950m, 910w, 860m. 'H-NMR (400 MHz, CDCl<sub>3</sub>): 5.79 (dd, J(H,P) = 10.6, J = 2.7, H–C(2)); 5.56 (d, J = 9.8, NH); 5.55 (dt, J = 8.2, 2.4, H–C(3)); 5.46 (ddd, J = 6.6, 2.8, 1.4, H–C(6)); 5.30 (dt, J = 3.0, 6.3, H–C(7)); 4.40 (dd, J = 12.5, 3.0, 1 H–C(8)); 4.39 (dd, J = 9.8, 3.0, H–C(5)); 4.35 (q, fine struct., J = 8.9, H–C(4)); 4.13 (dd, J = 12.4, 6.1, 1 H–C(8)); 3.82 (d, J(H,P) = 11.3, 3 H, P(OMe)<sub>2</sub>); 3.78 (d, J(H,P) = 11.2, 3 H, P(OMe)<sub>2</sub>); 2.13, 21.08, 2.07, 2.05 (d s, 4 AcO); 1.94 (s, (5.57, 113.56 (dd, J(C,P) = 23.1); 76.55 (dd, J(C,P) = 7.6); 69.72 (d); 68.22 (dd, J(C,P) = 15.5); 67.19 (d); 61.84 (f); 53.67 (dq, J(C,P) = 6.0); 53.33 (dq, J(C,P) = 5.8); 46.63 (d); 23.09 (q); 20.82 (q); 20.72 (2q); 20.66 (q). <sup>31</sup>P-NMR (81 MHz, CDCl<sub>3</sub>): 9.25. CI-MS: 524 (3,  $[M + 1]^*$ ), 464 (100).

4-Acetamido-2,4-dideoxy-D-glycero-D-galacto-oct-1-enopyranosyl)phosphonic Acid (10). A soln. of 35 (11.0 mg, 21 µmol) and CH<sub>2</sub>Cl<sub>2</sub> (1 ml) was cooled to 0°, and Me<sub>3</sub>SiBr (15 µl, 116 µmol, 5.5 equiv.) was added. After stirring for 24 h at 0°, MeOH (5 ml) was added and the soln. evaporated. The residue was dissolved in MeOH (1 ml) and 0.5M NaOMe/MeOH (168 µl, 4 equiv.) added. After stirring for 1 h at r.t., the soln. was filtered through *Dowex 50WX4* (H<sup>+</sup> form) and loaded on a *Dowex 1 × 8* column (HCOO<sup>-</sup> form, 10 ml). Elution with a HCOOH gradient (0→1.0M, 200 ml) gave, after freeze-drying, **10** (4.1 mg, 54%).  $R_t$  (PrOH/H<sub>2</sub>O 7:3) 0.28. 'H-NMR (400 MHz, D<sub>2</sub>O): 5.51 (dd, *J*(H,P) = 9.7, *J* = 2.2, H–C(2)); 4.46 (dt, *J* = 8.7, 2.2, H–C(3)); 4.23 (br. d, *J* = 10.7, H–C(5)); 4.10 (dd, *J* = 10.7, 9.1, H–C(4)); 3.95 (ddd, *J* = 9.8, 6.4, 2.6, H–C(7)); 3.92 (dd, *J* = 12.4, 2.5, H–C(8)); 3.68 (dd, *J* = 12.1, 6.6, H–C(8)); 3.63 (br. d, *J* = 9.8, H–C(6)); 2.10 (s, AcN). <sup>13</sup>C-NMR (100 MHz, D<sub>2</sub>O): (75.08 (s); 151.62 (d, *J*(C,P) = 212.6, C(1)); 110.65 (dd, *J*(C,P) = 22.4, C(2)); 76.12 (dd, *J*(C,P) = 9.2, C(5)); 70.12 (d); 68.61 (d); 67.84 (dd, *J*(C,P) = 13.5, C(3)); 63.52 (t, C(8)); 50.38 (d, C(4)); 22.37 (q). <sup>31</sup>P-NMR (162 MHz, D<sub>2</sub>O): 3.55.

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